Molecular genetics of gastroenteropancreatic neuroendocrine tumors

Lee F. Starker and Tobias Carling

Department of Surgery, Yale University School of Medicine, New Haven, Connecticut, USA

Correspondence to Tobias Carling, MD, PhD, Department of Surgery, Yale University School of Medicine, 330 Cedar Street, TMP202, New Haven, CT 06520, USA

Tel: +1 203 737 2036; fax: +1 203 737 4067; e-mail: tobias.carling@yale.edu

Current Opinion in Oncology 2008, 21:29-33

Purpose of review

Gastroenteropancreatic neuroendocrine tumors (GEP NETs) are relatively rare neoplasias arising from the embryonic neural crest, neuroectoderm and endoderm. GEP NETs occur either sporadically or as part of endocrine tumor susceptibility syndromes such as multiple endocrine neoplasia type 1 (MEN1), von Hippel Lindau (VHL) syndrome, neurofibromatosis (NF-1), and possibly tuberous sclerosis (TSC). The overall incidence of GEP NETs shows a significant increase over the past three decades. Improved understanding of the molecular genetics associated with these lesions will likely enhance the diagnosis and treatment of patients with GEP NET.

Recent findings

The molecular and clinical genetics of familial GEP NETs have been further elucidated by the characterization of the tumor suppressor genes, *MEN1*, *VHL*, *NF-1*, *TSC1*, and *TSC2*. The vastly improved technology in the field of cancer genetics with higher resolution of the study of genetic alterations, and the ability of unbiased mutational analyses of entire tumor genomes is likely to further the understanding of the genetic mechanisms of sporadic GEP NET as well.

Summary

Recent advances in the molecular genetics of sporadic and familial GEP NET are reviewed.

Keywords

carcinoid, endocrine, genetic, molecular, pancreas, tumor

Curr Opin Oncol 21:29-33 © 2008 Wolters Kluwer Health | Lippincott Williams & Wilkins 1040-8746

Introduction

Tumors that are derived from the diffuse neuroendocrine system of the gastrointestinal tract and pancreas are fairly rare. They occur in tissues that contain cells derived from embryonic neural crest, neuroectoderm, and endoderm. They, therefore, occur throughout the entirety of the body, albeit with a predilection for the lung and bronchus and the gastro-enter-pancreatic axis. Siegfrid Oberndorfer first coined the term karzinoide (carcinoid; cancerlike) in 1907. Gastroenteropancreatic neuroendocrine tumors (GEP NETs) include gastrointestinal carcinoids (also known as gastrointestinal NETs; GI NETs), as well as the neuroendocrine tumors of the pancreas (PNETs).

GEP NETs are relatively rare in contrast to adenocarcinomas, with an annual incidence of approximately 2.5 to five new cases per 100 000 [1]. However, a substantial increase in the incidence of these tumors has occurred over the last 30 years, with 460–720% increased prevalence [1]. Although the exact mechanism for this finding is unclear, advancements in technologies such as endoscopy and imaging techniques are likely to have contributed to the increased diagnosis of GEP NET. Although GEP NETs are increasingly being diagnosed, a concomitant improvement in outcomes has not been noted. The 5-year overall survival for all carcinoids and intestinal carcinoids in the United States between 1973 and 2002 has remained at 60% [1]. It should be pointed out, however, that there exists significant heterogeneity in outcomes between various groups of GEP NET. This is exemplified by 5-year survival rates of 97 and 30% for benign pancreatic insulinomas vs. nonfunctioning endocrine pancreatic tumors, respectively [2].

Familial gastroenteropancreatic neuroendocrine tumors

Although a majority of GEP NETs are sporadic, the molecular and clinical genetics of tumor susceptibility syndromes, in which GEP NETs may occur, have contributed to the understanding of tumorigenesis in these patients.

1040-8746 © 2008 Wolters Kluwer Health | Lippincott Williams & Wilkins

DOI:10.1097/CCO.0b013e328319ea7b

Copyright © Lippincott Williams & Wilkins. Unauthorized reproduction of this article is prohibited.

 Table
 1 Gastroenteropancreatic
 neuroendocrine
 tumors

 associated with MEN1 and their approximate penetrance

| Tumor type | Penetrance (%) |
|--|----------------|
| Enteropancreatic tumors | |
| Gastrinoma | 40 |
| Insulinoma | 10 |
| Nonfunctioning | 20 |
| Other (glucagonoma, VIPoma, somatostinoma) | 2 |
| Foregut carcinoids | |
| Gastric carcinoids | 10 |
| Thymic carcinoid | 2 |
| Bronchial carcinoid | 2 |

VIP, vasoactive intestinal peptide.

Multiple endocrine neoplasia type 1

Multiple endocrine neoplasia type 1 (MEN1; OMIM 131100) is an autosomal dominant endocrine tumor susceptibility syndrome causing tumors of the parathyroid, enteropancreatic endocrine tissue, and anterior pituitary [3]. MEN1 is relatively rare (approximately one in 30 000), and a consensus definition of MEN1 is used widely [4]. An MEN1 case has tumor in two of the three principal organs (parathyroid, enteropancreatic endocrine tissue, and anterior pituitary). Similarly, familial MEN1 is defined as one MEN1 case plus one first-degree relative with one of the three principal tumors. GEP NETs are the second most common manifestation of MEN1 after primary hyperparathyroidism and include a spectrum of tumors with variable penetrance in the disease (Table 1).

The *MEN1* gene, localized at chromosome 11q13, was identified by positional cloning [5,6]. Germline *MEN1* mutation is identifiable in 70–90% of typical MEN1 families, and some without an identified mutation may have large deletions or intron mutations not recognizable by polymerase chain reaction. The importance of *MEN1* gene inactivation in tumorigenesis is further substantiated by the fact that sporadic tumors, including GEP NETs, parathyroid adenomas, and bronchial carcinoid frequently harbor *MEN1* gene mutations [5–12]. The patterns of germline and somatic mutations in *MEN1* are similar, and approximately 80% predict truncation or absence of the encoded menin. There is no strong pattern of genotype/phenotype relations among germline and somatic *MEN1* mutations.

Menin is a 67-kDa protein, widely expressed, located in the nucleus, but also detected in cytoplasm and about telomeres [13]. Sequence analysis shows no homologous proteins. Through various protein interaction studies, menin has been suggested to partner with a number of other proteins such as transcription factors, DNA repair factors, cytoskeleton-associated proteins, among others [14–22]. Although the potential protein partners vary in function, most of them are nuclear proteins, generally involved in transcriptional regulation. The role of menin in transcriptional regulation was further substantiated as a large menin complex was demonstrated to exhibit lysine-4 histone methyltransferase activity, and that menin can directly bind to DNA [23,24]. Additionally, menin acts as a molecular adaptor linking the mixed-lineage leukemia (MLL) histone methyltransferase with lens epitheliumderived growth factor (LEDGF) [25[•]]. A recent study demonstrated that *MEN1*-causing missense mutations lead to a loss of function of menin due to enhanced proteolytic degradation, which may be a mechanism for inactivating menin as a tumor suppressor [26]. Heterozygous knockout of Men1 in mice provides an excellent model of MEN1 [27-29]. Mice develop normally, but by 16 months frequently develop parathyroid tumors, insulinomas, and prolactinomas. Interestingly, insulinomas in these mice can develop in the absence of chromosome instability or microsatellite instability [30]. In a recent study on MEN1 patients, however, Perren et al. [31[•]] found that loss of heterozygosity (LOH) of the MEN1 locus in all 27 endocrine pancreatic microadenomas and 19 of 20 (95%) monohormonal endocrine cell clusters.

von Hippel Lindau syndrome

The von Hippel Lindau (VHL; OMIM 193300) disease is an autosomal dominant neoplasia syndrome that results from germline mutations in the VHL gene [32]. These mutations lead to the development of several benign or malignant tumors and cysts in many organs. VHL is characterized by predisposition to develop hemangioblastomas of the retina and central nervous system (CNS), renal cell carcinomas and renal cysts, pheochromocytomas, endolymphatic sac tumors, as well as pancreatic lesions with marked phenotypic variability. Although cysts are the most common lesion (33–70%) in the pancreas in patients with VHL, endocrine pancreatic tumors occur in 11–17%, and have malignant potential [33].

The VHL gene is a tumor suppressor gene on the short arm of chromosome 3(3p25-26), with three exons encoding the VHL protein [34]. The VHL protein shuttles between the nucleus and cytoplasm, binding to elongin C, elongin B, Cul2, and Rbx1, and degrades alpha subunits of hypoxia-inducible factor in an oxygen-dependent manner [35]. Lack of degradation of this factor due to absence of the VHL protein results in uncontrolled production of factors promoting formation of blood vessels such as vascular endothelial growth factor implicated in tumor development [35]. Germline mutations in the VHL gene are extremely heterogeneous and are distributed widely throughout the coding sequence [32]. They are now identifiable in virtually all families with VHL [32]. The exact molecular mechanism of GEP NET development in VHL is unknown. Analyses of allelic loss identify genetic loci distinct from and mapping proximal to VHL within human chromosome 3p VHL kindred under study. Furthermore, chromosome 3p LOH occurs subsequent to VHL mutation and cyst formation and correlates with malignant progression in VHLassociated GEP NET. These findings suggest that additional genetic alterations, possibly in a stepwise fashion, cause GEP NET in the VHL syndrome [36].

Neurofibromatosis type 1 and tuberous sclerosis

Neurofibromatosis type 1 (NF-1; OMIM 162200) and tuberous sclerosis (TS; OMIM 191100) are both rare autosomal dominant tumor susceptibility syndromes. Ampullary carcinoids, duodenal and pancreatic somatostatinomas, and nonfunctioning GEP NETs have occasionally been reported in such kindreds [37,38].

NF-1 and TS are caused by inactivating mutations in the tumor suppressor genes *NF1* (17q11.2), *TSC1* (9q34), and *TSC2* (16p13.3), respectively. *NF1* encodes the protein neurofibromin, which also regulates *TSC1* and *TSC2* through the mammalian target of rapamycin (mTOR). Loss of function of the *NF1* gene causes mTOR activation and tumor development [39]. Interestingly, disruption of TSC2 in pancreatic beta cells induces beta cell mass expansion in an mTOR-dependent manner [40].

Sporadic gastroenteropancreatic neuroendocrine tumors

Through the clinical and molecular genetic studies of kindreds with GEP NETs, the underlying alterations in these families have been characterized. In contrast, less is known about the genetic mechanism of sporadic GEP NETs, although genes involved in their familial counterparts also play a role in the molecular pathology of sporadic tumors. There exist both similarities and differences in the tumorigenesis of various types of GEP NETs, and is here discussed in two broad groups, those that arise in the pancreas (PNETs) and the gastrointestinal tract (GI NETs) [41].

Sporadic endocrine pancreatic tumors (neuroendocrine tumors of the pancreas)

The occurrence of chromosomal gains, or losses, or both have been extensively studied in PNETs, by LOH analysis, comparative genomic hybridization (CGH), and array CGH analyses. Allelic loss is most commonly seen at chromosome loci 1p (23–75%), 1q (20–88%), 3p (25–62%), 11p (29–52%), 11q (28–66%), and 22q (38–93%), although many other loci throughout the genome show LOH in more than 20% of the tumors [10–12,42–44]. As noted, LOH at 11q13 is common, and *MEN1* gene mutations have been identified in sporadic gastrinomas, insulinomas, glucagonomas, VIPomas, and nonfunctioning tumors. The overall incidence of *MEN1* gene mutations in sporadic PNETs varies between 13 and 38% [11,12,43,44]. In contrast, the *VHL* gene seems not to be mutated in sporadic PNETs [45]. *DPC4/SMAD4* mutations are seen in about 20% of sporadic PNETs [46]. Deletions on chromosome 1, 3p, 6, 11q, 17p, 21q, and 22q as well as gains on chromosomes 4, 7, 14q, and Xq have been associated with malignancy in sporadic PETs [47]. Additional attempts to distinguish benign from malignant PNETs used genomewide expression microarray studies revealing more than three-fold overexpression of 66 transcripts including IGFBP3, which is deregulated in many tumor types. Underexpression (>three-fold) was seen for 119 transcripts, including p21, O6-MGMT, and JunD [48]. When comparing benign and malignant PETs, O6-MGMT was downregulated in malignant tumors and the protooncogene MET was upregulated together with IGFBP1 and IGFBP3 [49]. More recently, Duerr et al. [41] using a DNA microarray analysis and hierarchical clustering of 19 PNETs revealed a 'benign' and 'malignant' cluster. FEV, adenylate cyclase 2 (ADCY2), nuclear receptor subfamily 4, group A, member 2 (NR4A2), and growth arrest and DNA-damage-inducible, beta (GADD45b) were the most highly upregulated genes in the malignant group of PNETs.

Sporadic gastrointestinal carcinoids (gastroenteropancreatic neuroendocrine tumors)

Similar to PNETs, chromosomal gains, or losses, or both have been studied in GI NETs using LOH analysis, CGH, and array CGH analyses. Loss of chromosome 18 and, to a lesser extent, loss of chromosome loci 9p and 16q are the most common genetic alterations of GI NETs. Amplification of chromosomal loci is less common in GI NETs than in PNETs and other endocrine tumors, but occurs most frequently on chromosomes 4, 5, 7, 14, 17, and 20 [50°,51–56]. Interestingly, recent findings of genetic alterations in GI NETs using high-resolution array-based CGH analysis are similar to earlier studies using conventional CGH analysis [50°].

Conclusion

Despite the improved understanding of the molecular genetics of familial endocrine neoplasia syndromes, the exact mechanism of GEP NET development is still unclear. With improved technology in the field of cancer genetics, such as large-scale sequencing of entire tumor genomes [57,58], it is likely that significant advancements will occur in the understanding of GEP NET, as well. Further studies of gene function may lead to development of novel therapeutic modalities.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

Additional references related to this topic can also be found in the Current World Literature section in this issue (pp. 88-90).

 Modlin IM, Oberg K, Chung DC, et al. Gastroenteropancreatic neuroendocrine tumours. Lancet Oncol 2008; 9:61–72.

Copyright © Lippincott Williams & Wilkins. Unauthorized reproduction of this article is prohibited.

of special interest

of outstanding interest

32 Endocrine tumors

- 2 Oberg K, Eriksson B. Endocrine tumours of the pancreas. Best Pract Res Clin Gastroenterol 2005; 19:753–781.
- 3 Carling T. Multiple endocrine neoplasia syndrome: genetic basis for clinical management. Curr Opin Oncol 2005; 17:7–12.
- 4 Brandi ML, Gagel RF, Angeli A, et al. Guidelines for diagnosis and therapy of MEN type 1 and type 2. J Clin Endocrinol Metab 2001; 86:5658–5671.
- 5 Chandrasekharappa S, Guru S, Manickam P, et al. Positional cloning of the gene for multiple endocrine neoplasia type 1. Science 1997; 276:404–407.
- 6 The European Consortium, et al. Identification of the multiple endocrine neoplasia type 1 (MEN1) gene. Hum Mol Genet 1997; 6:1177–1183.
- 7 Carling T, Correa P, Hessman O, et al. Parathyroid MEN1 gene mutations in relation to clinical characteristics of nonfamilial primary hyperparathyroidism. J Clin Endocrinol Metab 1998; 83:2960–2963.
- 8 Farnebo F, Teh B-T, Kytölä S, et al. Alterations of the MEN1 gene in sporadic parathyroid tumors. J Clin Endocrinol Metab 1998; 83:2627–2630.
- 9 Heppner C, Kester MB, Agarwal SK, et al. Somatic mutations of the MEN1 gene in parathyroid tumours. Nat Genet 1997; 16:375–378.
- 10 Hessman O, Lindberg D, Einarsson A, et al. Genetic alterations on 3p, 11q13, and 18q in nonfamilial and MEN 1-associated pancreatic endocrine tumors. Genes Chromosomes Cancer 1999; 26:258–264.
- 11 Hessman O, Lindberg D, Skogseid B, et al. Mutation of the multiple endocrine neoplasia type 1 gene in nonfamilial, malignant tumors of the endocrine pancreas. Cancer Res 1998; 58:377–379.
- 12 Hessman O, Skogseid B, Westin G, Akerstrom G. Multiple allelic deletions and intratumoral genetic heterogeneity in men1 pancreatic tumors. J Clin Endocrinol Metab 2001; 86:1355–1361.
- 13 Agarwal SK, Lee Burns A, Sukhodolets KE, *et al.* Molecular pathology of the MEN1 gene. Ann N Y Acad Sci 2004; 1014:189–198.
- 14 Agarwal S, Guru S, Heppner C, et al. Menin interacts with the AP1 transcription factor JunD and represses JunD-activated transcription. Cell 1999; 96:143–152.
- 15 Busygina V, Suphapeetiporn K, Marek LR, et al. Hypermutability in a Drosophila model for multiple endocrine neoplasia type 1. Hum Mol Genet 2004; 13:2399-2408.
- 16 Jin S, Mao H, Schnepp RW, et al. Menin associates with FANCD2, a protein involved in repair of DNA damage. Cancer Res 2003; 63:4204–4210.
- 17 Kim H, Lee JE, Cho EJ, et al. Menin, a tumor suppressor, represses JunDmediated transcriptional activity by association with an mSin3A-histone deacetylase complex. Cancer Res 2003; 63:6135–6139.
- 18 Schnepp RW, Hou Z, Wang H, et al. Functional interaction between tumor suppressor menin and activator of S-phase kinase. Cancer Res 2004; 64:6791-6796.
- 19 Schnepp RW, Mao H, Sykes SM, et al. Menin induces apoptosis in murine embryonic fibroblasts. J Biol Chem 2004; 279:10685–10691.
- 20 Sowa H, Kaji H, Hendy GN, et al. Menin is required for bone morphogenetic protein 2- and transforming growth factor beta-regulated osteoblastic differentiation through interaction with Smads and Runx2. J Biol Chem 2004; 279:40267-40275.
- 21 Sowa H, Kaji H, Kitazawa R, et al. Menin inactivation leads to loss of transforming growth factor beta inhibition of parathyroid cell proliferation and parathyroid hormone secretion. Cancer Res 2004; 64:2222-2228.
- 22 Stalberg P, Grimfjard P, Santesson M, et al. Transfection of the multiple endocrine neoplasia type 1 gene to a human endocrine pancreatic tumor cell line inhibits cell growth and affects expression of JunD, delta-like protein 1/ preadipocyte factor-1, proliferating cell nuclear antigen, and QM/Jif-1. J Clin Endocrinol Metab 2004; 89:2326-2337.
- 23 Hughes CM, Rozenblatt-Rosen O, Milne TA, et al. Menin associates with a trithorax family histone methyltransferase complex and with the hoxc8 locus. Mol Cell 2004; 13:587–597.
- 24 La P, Silva AC, Hou Z, et al. Direct binding of DNA by tumor suppressor menin. J Biol Chem 2004; 19:49045-49054.
- Yokoyama A, Cleary ML. Menin critically links MLL proteins with LEDGF on
 cancer-associated target genes. Cancer Cell 2008; 14:36–46.

This elegant study further establishes the dual role of menin as a suppressor in endocrine tumorigenesis and as an essential oncogenic cofactor in leukemic transformation.

- 26 Yaguchi H, Ohkura N, Takahashi M, et al. Menin missense mutants associated with multiple endocrine neoplasia type 1 are rapidly degraded via the ubiquitin-proteasome pathway. Mol Cell Biol 2004; 24:6569-6580.
- 27 Bertolino P, Tong WM, Galendo D, et al. Heterozygous Men1 mutant mice develop a range of endocrine tumors mimicking multiple endocrine neoplasia type 1. Mol Endocrinol 2003; 17:1880–1892.

- 28 Biondi CA, Gartside MG, Waring P, et al. Conditional inactivation of the MEN1 gene leads to pancreatic and pituitary tumorigenesis but does not affect normal development of these tissues. Mol Cell Biol 2004; 24:3125– 3131.
- 29 Crabtree JS, Scacheri PC, Ward JM, et al. Of mice and MEN1: insulinomas in a conditional mouse knockout. Mol Cell Biol 2003; 23:6075–6085.
- 30 Scacheri PC, Kennedy AL, Chin K, et al. Pancreatic insulinomas in multiple endocrine neoplasia, type I knockout mice can develop in the absence of chromosome instability or microsatellite instability. Cancer Res 2004; 64:7039-7044.
- 31 Perren A, Anlauf M, Henopp T, et al. Multiple endocrine neoplasia type 1
- (MEN1): loss of one MEN1 allele in tumors and monohormonal endocrine cell clusters but not in islet hyperplasia of the pancreas. J Clin Endocrinol Metab 2007; 92:1118–1128.

This study identified allelic loss at the *MEN1* gene locus in small microadenomas and monohormonal endocrine cell clusters in the pancreas of patients with MEN1, suggesting that such genetic abnormalities are early events and contribute to the development of the classic occurrence of multiple small endocrine pancreatic tumors in these patients.

- 32 Lonser RR, Glenn GM, Walther M, et al. von Hippel-Lindau disease. Lancet 2003; 361:2059–2067.
- 33 Corcos O, Couvelard A, Giraud S, et al. Endocrine pancreatic tumors in von Hippel-Lindau disease: clinical, histological, and genetic features. Pancreas 2008; 37:85–93.
- 34 Latif F, Tory K, Gnarra J, et al. Identification of the von Hippel-Lindau disease tumor suppressor gene. Science 1993; 260:1317–1320.
- 35 Kaelin WG Jr. Molecular basis of the VHL hereditary cancer syndrome. Nat Rev Cancer 2002; 2:673-682.
- 36 Lott ST, Chandler DS, Curley SA, et al. High frequency loss of heterozygosity in von Hippel-Lindau (VHL)-associated and sporadic pancreatic islet cell tumors: evidence for a stepwise mechanism for malignant conversion in VHL tumorigenesis. Cancer Res 2002; 62:1952–1955.
- 37 Eledrisi MS, Stuart CA, Alshanti M. Insulinoma in a patient with tuberous sclerosis: is there an association? Endocr Pract 2002; 8:109-112.
- 38 Garbrecht N, Anlauf M, Schmitt A, et al. Somatostatin-producing neuroendocrine tumors of the duodenum and pancreas: incidence, types, biological behavior, association with inherited syndromes, and functional activity. Endocr Relat Cancer 2008; 15:229–241.
- 39 Johannessen CM, Reczek EE, James MF, et al. The NF1 tumor suppressor critically regulates TSC2 and mTOR. Proc Natl Acad Sci U S A 2005; 102:8573–8578.
- 40 Rachdi L, Balcazar N, Osorio-Duque F, et al. Disruption of Tsc2 in pancreatic beta cells induces beta cell mass expansion and improved glucose tolerance in a TORC1-dependent manner. Proc Natl Acad Sci U S A 2008; 105:9250– 9255.
- 41 Duerr EM, Mizukami Y, Ng A, et al. Defining molecular classifications and targets in gastroenteropancreatic neuroendocrine tumors through DNA microarray analysis. Endocr Relat Cancer 2008; 15:243–256.
- 42 Lindberg D, Akerstrom G, Westin G. Mutational analyses of WNT7A and HDAC11 as candidate tumour suppressor genes in sporadic malignant pancreatic endocrine tumours. Clin Endocrinol (Oxf) 2007; 66:110– 114.
- 43 Wang EH, Ebrahimi SA, Wu AY, et al. Mutation of the MENIN gene in sporadic pancreatic endocrine tumors. Cancer Res 1998; 58:4417– 4420.
- 44 Zhuang Z, Vortmeyer AO, Pack S, et al. Somatic mutations of the MEN1 tumor suppressor gene in sporadic gastrinomas and insulinomas. Cancer Res 1997; 57:4682–4686.
- 45 Chung D, Smith A, Louis D, et al. A novel pancreatic endocrine tumor suppressor gene locus on chromosome 3p with clinical prognostic implications. J Clin Invest 1997; 100:404-410.
- 46 Bartsch D, Hahn SA, Danichevski KD, et al. Mutations of the DPC4/Smad4 gene in neuroendocrine pancreatic tumors. Oncogene 1999; 18:2367– 2371.
- 47 Toumpanakis CG, Caplin ME. Molecular genetics of gastroenteropancreatic neuroendocrine tumors. Am J Gastroenterol 2008; 103:729-732.
- 48 Maitra A, Hansel DE, Argani P, et al. Global expression analysis of well differentiated pancreatic endocrine neoplasms using oligonucleotide microarrays. Clin Cancer Res 2003; 9:5988–5995.
- 49 Hansel DE, Rahman A, House M, et al. Met proto-oncogene and insulin-like growth factor binding protein 3 overexpression correlates with metastatic ability in well differentiated pancreatic endocrine neoplasms. Clin Cancer Res 2004; 10:6152–6158.

 Kulke MH, Freed E, Chiang DY, et al. High-resolution analysis of genetic
 alterations in small bowel carcinoid tumors reveals areas of recurrent amplification and loss. Genes Chromosomes Cancer 2008; 47:591– 603.

Using array CGH, this study provides a basis for further investigation of putative oncogenes and tumor suppressor genes involved in the development of GI NETs.

- 51 Kytola S, Hoog A, Nord B, et al. Comparative genomic hybridization identifies loss of 18q22-qter as an early and specific event in tumorigenesis of midgut carcinoids. Am J Pathol 2001; 158:1803–1808.
- 52 Lollgen RM, Hessman O, Szabo E, et al. Chromosome 18 deletions are common events in classical midgut carcinoid tumors. Int J Cancer 2001; 92:812–815.
- 53 Terris B, Meddeb M, Marchio A, et al. Comparative genomic hybridization analysis of sporadic neuroendocrine tumors of the digestive system. Genes Chromosomes Cancer 1998; 22:50–56.

- 54 Wang GG, Yao JC, Worah S, et al. Comparison of genetic alterations in neuroendocrine tumors: frequent loss of chromosome 18 in ileal carcinoid tumors. Mod Pathol 2005; 18:1079–1087.
- 55 Zhao J, de Krijger RR, Meier D, et al. Genomic alterations in well differentiated gastrointestinal and bronchial neuroendocrine tumors (carcinoids): marked differences indicating diversity in molecular pathogenesis. Am J Pathol 2000; 157:1431–1438.
- 56 Zikusoka MN, Kidd M, Eick G, et al. The molecular genetics of gastroenteropancreatic neuroendocrine tumors. Cancer 2005; 104:2292–2309.
- 57 Sjoblom T. Systematic analyses of the cancer genome: lessons learned from sequencing most of the annotated human protein-coding genes. Curr Opin Oncol 2008; 20:66–71.
- 58 Sjoblom T, Jones S, Wood LD, et al. The consensus coding sequences of human breast and colorectal cancers. Science 2006; 314:268–274.